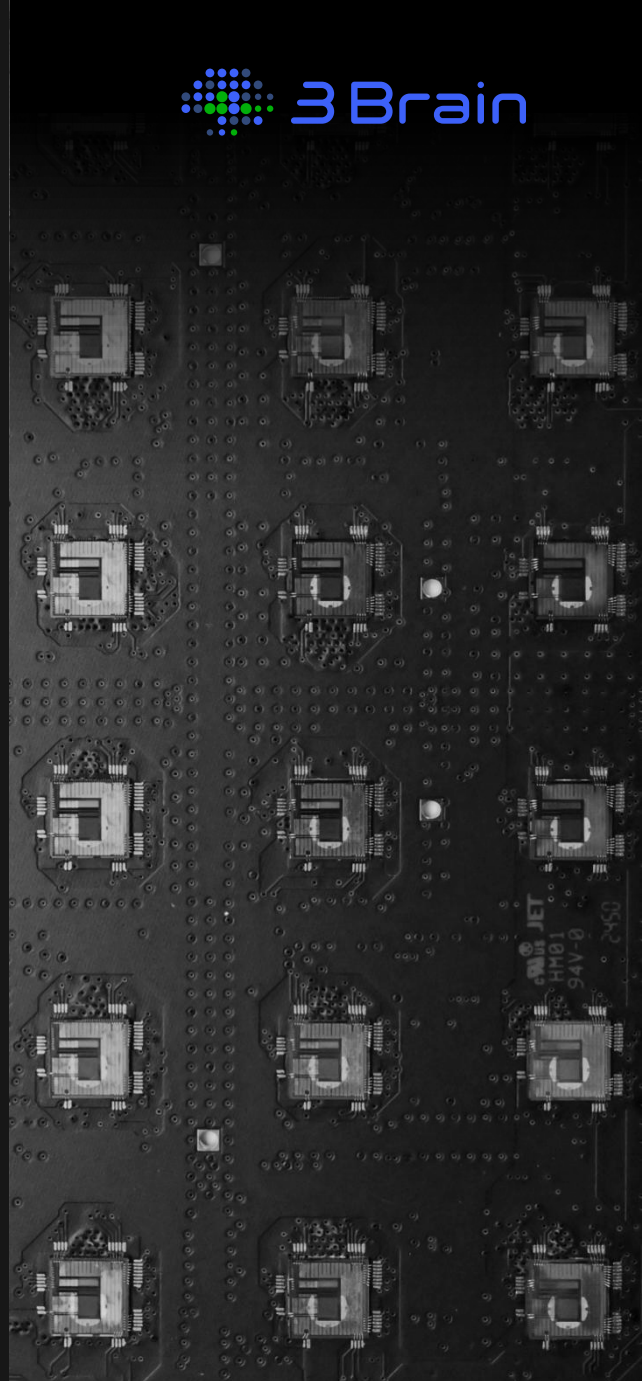


CorePlate™ 6W Handling, Cleaning, Sterilization & Hydrophilization Guide

Last update: 01 Apr 25
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CorePlate™ 6W General Handling

CorePlate™ 6W is an integrated low-power active electronic circuit. Proper handling is essential to ensure optimal function and prevention of damage.

CorePlate™ 6W refers to the following:

- CorePlate™ 6W 38/60

General Handling:

- As a general rule only the wells should be in contact with liquid while the rest of the plate should remain dry. Exception: for ethanol sterilization, the gaps between the wells may be filled with 70% ethanol as described in “Sterilization Procedure”.
- Do not touch the contact pads on the bottom of the plate (see Fig. 1). Always handle CorePlate™ 6W by using plastic gloves.
- Do not place in direct contact with a metallic worktop. Use a plastic tray below CorePlate™ 6W when in an incubator, or under a hood. Electrostatic charges on metallic surfaces may damage the on-chip circuits and result in damaging the device.
- Do not touch the Active Area (see Fig. 1) of the microchip integrated in each well of the CorePlate™ 6W with any tools except those specifically allowed (see section “Cleaning Procedure for CorePlate™ 6W”). In particular, hard or semi-hard objects (e.g. metal or plastic tools) can irreparably damage the electrodes and the on-chip circuits.
- Do not autoclave CorePlate™ 6W. For CorePlate™ 6W sterilization, please refer to the sterilization procedure.
- Maintain the pH of the electrophysiological solutions used for neuronal cultures or brain tissues, possibly at physiological conditions (pH 7-7.5). Strong changes in the pH of the solution might damage the electrodes.

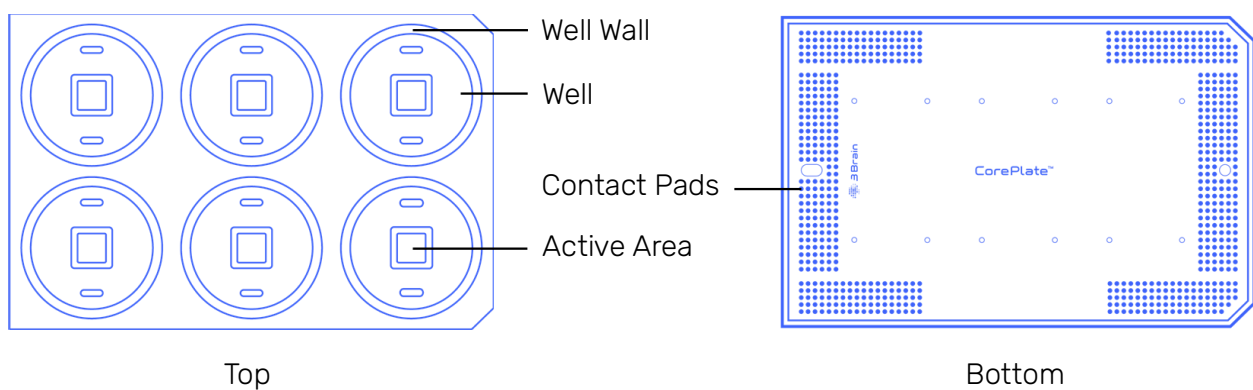


Figure 1: Diagram of the top and bottom of CorePlate™ 6W.

Precautions to follow during recording:

- Before inserting CorePlate™ 6W into the HyperCAM Alpha, clean the contact pads on the bottom of CorePlate™ 6W (Fig. 1) with tissue paper soaked in ethanol 96% and let it dry for a few seconds.
- Prevent liquids from spilling out of CorePlate™ 6W into the HyperCAM Alpha.
- In order to ensure a good quality of the recordings please remember that CorePlate™ 6W are light-sensitive devices and their performances might be affected by high-light intensity.

Cleaning procedure for CorePlate™ 6W

CorePlate™ 6W are warranted as single-use plates.

If re-use is necessary, CorePlate™ 6W must be thoroughly cleaned following the proper steps indicated to maintain performance and prevent contamination.

General Cleaning Guidelines:

Prevention of Damage. During each step of this procedure, ensure the pipette tip (or any other object) does not directly touch the Active Area.

Avoid Wetting the Contact Pads. While cleaning CorePlate™ 6W, it is highly recommended not to wet the contact pads on the bottom of the plate (see Fig. 1).

Standard Cleaning Procedure:

1. **Rinse Wells.** Rinse the wells of CorePlate™ 6W thoroughly with Double Deionized Water (ddH₂O).
2. **Detergent Application.** Remove the ddH₂O and fill the wells with a detergent such as 5% Extran (Merck), 1% Terg-A-zyme (Alconox) or 1% SDS, and gently pipette for a few seconds.
3. **Soak.** Leave the detergent for a few minutes (typically 3-5 min) and then pipette again to dislodge any remaining residue.
4. **Final Rinse.** Remove the detergent and rinse the wells of CorePlate™ 6W thoroughly with ddH₂O, then leave the wells filled with ddH₂O for 1-2 minutes and repeat this operation 3-4 times in order to be assured of washing out the detergent completely.

Intense Cleaning Procedure:

If the chips of CorePlate™ 6W remain dirty after the standard cleaning procedure, you may have to repeat this procedure, including gently brushing the surface of each Active Area (see Fig. 1) with a soft, clean paintbrush during the Detergent Application step. Be aware that this operation can damage the Active Area of the chip and is advised only as a final option if the standard cleaning procedure fails to fully clean the chip. Ensure that the paintbrush is completely clean to avoid introducing more debris into the environment, and ensure that brushing is performed very gently and with utmost care to minimize the risk of damage to the electrodes.

Drying and Storage:

Drying. CorePlate™ 6W can be dried by using a gentle flux of Nitrogen air (do not expose the active area to an intense flux to avoid potential electrode damage) or under a biological hood. Alternatively, let CorePlate™ 6W dry on a bench, making sure to cover the wells to avoid dust deposition inside the wells.

Cleaning the Exterior. The area out of the wells of CorePlate™ 6W can be cleaned with a tissue soaked in ethanol >96%. Ensure that gloves are worn whilst handling the plate.

Storage. Once CorePlate™ 6W is dried it should be stored closed (with the lid on) in a box in order to protect them from dust and other contaminants.

Sterilization procedure for CorePlate™ 6W

As indicated in the previous section, CorePlate™ 6W are warranted as single-use plates.

Sterilization may be required before use (e.g. cell culturing). In this case, we recommend UV-light sterilization as the preferred method, and in the case this is not available, EtOH Sterilization may also be used.

General Sterilization Guidelines:

Prevention of Damage. During each step of this procedure, ensure the pipette tip (or any other object) does not directly touch the Active Area.

UV-light Sterilization:

Expose CorePlate™ 6W with the lid open to UVC-light for 45 min (dependant on the UV-light intensity and power).

Ensure that the light is well illuminating the wells in a homogeneous way, ideally the light should come from directly above to guarantee a strong and consistent UV-illumination of the wells.

The same procedure applies to sterilize the lid of CorePlate™ 6W.

EtOH Sterilization:

Perform the following steps under a biological hood:

1. **70% EtOH Application.** Fill the Wells of CorePlate™ 6W completely with 70% ethanol up to the rim of the Well Wall (Fig. 2). Also, thoroughly wet the area in between the wells with 70% EtOH ensuring that all edges are covered with EtOH.
2. **70% EtOH Soak.** Wait for 30-45 minutes to allow for sterilization.
3. **EtOH Removal.** Remove the ethanol using sterile tips and proper sterile technique.
4. **Rinse.** Fill $\frac{3}{4}$ of the wells of CorePlate™ 6W with sterile ddH₂O, wait a few seconds and remove the liquid with a sterile tip. Repeat this step in order to ensure the complete washing out of ethanol.
5. **Dry.** When removing sterile ddH₂O for the last time, be sure to completely dry the wells. Carefully aspirate any remaining liquid from the recording area, keeping the pipette tip close to the well/chip border but not touching the chip to avoid damaging the electrodes.

EtOH sterilization may also be done in combination with UV-light sterilization.

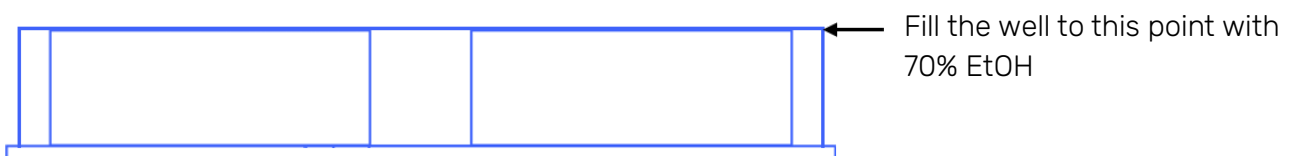


Figure 2: Side view diagram of CorePlate™ 6W showing the fill level of the wells.

Hydrophilization procedure for CorePlate™ 6W

In the case that CorePlate™ 6W displays hydrophobic properties (e.g. after a long period of disuse), perform the following hydrophilization protocol.

General Hydrophilization Guidelines:

Prevention of Damage. During each step of this procedure, ensure the pipette tip (or any other object) does not directly touch the Active Area.

Avoid Wetting the Contact Pads. While hydrophilizing CorePlate™ 6W, it is highly recommended not to wet the contact pads on the bottom of the plate (see Fig. 1).

Standard Hydrophilization Procedure:

1. **Rinse Wells with 70% EtOH.** Rinse the wells of CorePlate™ 6W (Fig. 1) thoroughly with 70% EtOH and delicately flush the liquid on each Active Area with a Pasteur pipette.
2. **Gently Brush Wells.** Gently brush each Active Area with a paintbrush. Ensure that the paintbrush is completely clean to avoid introducing debris into the environment, and ensure that brushing is performed very gently and with utmost care to minimize the risk of damage to the electrodes.
3. **Rinse Wells With ddH₂O.** Remove the 70% EtOH and rinse the wells of CorePlate™ 6W thoroughly with ddH₂O. Delicately flush ddH₂O on each Active Area with a Pasteur pipette.
4. **Gently Brush Wells.** Gently brush the Active Area with a paintbrush. Ensure that the paintbrush is completely clean to avoid introducing debris into the environment, and ensure that brushing is performed very gently and with utmost care to minimize the risk of damage to the electrodes.
5. **Rinse Wells With Phosphate Buffered Saline (PBS).** Remove the ddH₂O and rinse the wells of CorePlate™ 6W thoroughly with PBS. Delicately flush PBS on each Active Area with a Pasteur pipette.
6. **Soak Wells in PBS.** After rinsing with PBS, fill the wells with PBS. Leave to soak for 1-2 minutes.

If the chips remain hydrophobic, repeat the procedure a couple of times. If necessary, leave the chips soaked in PBS overnight. If the problem persists, please contact our customer success team at cs@3brain.com.